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EXAMINER

KUBELIK, ANNE R

ART UNIT	PAPER NUMBER
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1638

DATE MAILED: 05/18/2006

Please find below and/or attached an Office communication concerning this application or proceeding.



### **DETAILED ACTION**

1. Prosecution in this application has been reopened in light of the modified rejections below.
2. The text of those sections of Title 35, U.S. Code not included in this action can be found in a prior Office action.

#### ***Claim Rejections - 35 USC § 112***

3. Claims 1-3, 9-12, 17-19, 38, 43, 46, 49, 52 and 55-64 are rejected under 35 U.S.C. 112, first paragraph, because the specification, while being enabling for nucleic acids encoding SEQ ID NO:2 and 10, expression cassettes comprising the nucleic acids, plants and seeds comprising a construct comprising the nucleic acid, and a method of using it to impact a plant pest, does not reasonably provide enablement for any nucleic acid that has 90% identity to SEQ ID NO:1, expression cassettes comprising the nucleic acid, plants and seeds comprising a construct comprising the nucleic acid and a method of using it to impact a plant pest. The specification does not enable any person skilled in the art to which it pertains, or with which it is most nearly connected, to make and/or use the invention commensurate in scope with these claims. The rejection is modified from the rejection set forth in the Office action mailed 6 May 2005. Applicant's arguments filed 23 February 2006 in a Reply Brief have been fully considered but they are not persuasive.

The claims are broadly drawn to any nucleic acid that has 90%, 93%, 94% or 95% identity to SEQ ID NO:1, expression cassettes comprising the nucleic acids, plants and seeds

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comprising constructs comprising the nucleic acids, and a method of using the constructs to impact a plant pest.

The instant specification, however, only provides guidance for methods of assaying the activity of *B. thuringiensis* strain 1218 and lysate against Western corn rootworm and Southern corn rootworm (examples 1 and 2); isolation of crystal protein from the strain and assaying of it for pesticidal activity against western corn rootworm (example 3); identification of two coding regions, *Cry1218-1* and *Cry1218-2* (SEQ ID NO:1 and 3, with SEQ ID NOs:27 and 28 as the genomic clones), isolated by unknown methods, as encoding proteins, SEQ ID NOs:2 and 4, respectively, that have homology to Cry8Ba1 (example 4); production in *E. coli* of truncated proteins, SEQ ID NOs:15 and 17, encoded by SEQ ID NOs:16 and 18, respectively, that are active against southern corn rootworm (example 4); and production of maize-preferred coding sequences of a different truncated version of Cry1218-1 - the nucleic acid is SEQ ID NO:9, which encodes SEQ ID NO:10 (example 5). The specification also teaches making two mutant versions of truncated Cry1218-1 (SEQ ID NO:16), one of which has a truncated N-terminus (comprising amino acids 43-663 of SEQ ID NO:16), and the other in which the 4 amino acid sequence NGSR has been inserted after amino acid 164 - all of these mutant proteins are effective against Colorado potato beetle (example 6) and other mutant proteins (SEQ ID NOs:32, 34, 42 and 46) that have added chymotrypsin cleavage sites - all are more effective against southern and western corn rootworm than Cry1218-1 (example 7). The specification also teaches transformation of maize with SEQ ID NO:9 (examples 8 and 9).

The instant specification fails to provide guidance for nucleic acid that has 90%, 93%, 94% or 95% identity to the full-length of SEQ ID NO:1, expression cassettes comprising the

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nucleic acid, plants and seeds comprising a construct comprising the nucleic acid and a method of using it to impact a plant pest.

The instant specification fails to provide guidance for which amino acids of SEQ ID NO:2 can be altered and to which other amino acids, and which amino acids must not be changed, to maintain Cry8 activity of the encoded protein. The specification also fails to provide guidance for which amino acids can be deleted and which regions of the protein can tolerate insertions and still produce a functional enzyme.

The specification on pg 28, lines 5-16, suggests making these nucleic acids by making conservative substitutions in the encoded protein. However, making “conservative” substitutions (*e.g.*, substituting one polar amino acid for another, or one acidic one for another) does not produce predictable results. Lazar et al (1988, Mol. Cell. Biol. 8:1247-1252) showed that the “conservative” substitution of glutamic acid for aspartic acid at position 47 reduced biological function of transforming growth factor alpha while “nonconservative” substitutions with alanine or asparagine had no effect (abstract). Similarly, Hill et al (1998, Biochem. Biophys. Res. Comm. 244:573-577) teach that when three histidines that are maintained in ADP-glucose pyrophosphorylase across several species are substituted with the “nonconservative” amino acid glutamine, there is little effect on enzyme activity, while the substitution of one of those histidines with the “conservative” amino acid arginine drastically reduced enzyme activity (see Table 1). The nucleic acids encoding all these mutated proteins, however, would have much greater than 90% identity to the nucleic acids encoding the original protein.

Making amino acid substitutions in *cry* proteins is unpredictable. Each *cry* protein only has activity against one or few insect species (de Maagd et al, 1999, Appl. Environ. Microbiol.

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65:4369-4374, see pg 4369, column 1, paragraph 1). Even a single amino acid substitution in a *cry* protein may alter its insecticidal specificity, and toxicity must be determined empirically (Tounsi et al, 2003, J. Appl. Microbiol. 95:23-28; see pg 27, column 2, paragraph 2). For example, a conservative substitution of a lysine for an arginine in a cry11A protein eliminated toxicity to *Aedes aegyptii* but not other pests to which the wild-type protein has toxicity (Angsuthanasombat et al, 2001, J. Biochem. Mol. Biol. 34:402-407, paragraph spanning the columns on pg 405).

The specification, on pg 65, lines 12-14, indicates that the instant SEQ ID NO:1 has homology to GenBank U04365, which is identical to SEQ ID NO:3 of Michaels et al (1996, US Patent 5,554,534). This nucleotide sequence has 85.1% identity to SEQ ID NO:1. The specification, on pg 29, suggests using GenBank U04365 (SEQ ID NO:3 of Michaels et al ) as the other nucleic acid in shuffling to create the claimed nucleic acids; however, it encodes a protein with 79.8% identity to the instant SEQ ID NO:2 (see sequence search report). Thus, this sequence cannot be used as guidance for nucleic acids with 90% identity to SEQ ID NO:1 and that encode proteins with 70% identity to SEQ ID NO:2, as encompassed by the full scope of the claims.

Furthermore, SEQ ID NO:3 of Michaels et al is pesticidal for *Cyclocephala borealis* (northern masked chafer) and *Popillia japonica* (Japanese beetle)(column 14, line 46, to column 15, line 2), whereas the instant SEQ ID NO:1 is pesticidal to *Diabrotica longicornis howardi* (southern corn rootworm). Given the teachings of Tounsi et al, de Maagd et al, and Angsuthanasombat et al, one of skill in the art could not even predict for which pests the protein encoded by nucleic acids that have 90% identity to SEQ ID NO:1 would be toxic, and the

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specification does not teach which amino acid substitutions must can be made to retain southern corn rootworm pesticidal activity.

Given the claim breath, unpredictability, and lack of guidance as discussed above, undue experimentation would have been required by one skilled in the art to develop and evaluate nucleic acids with 90% identity to SEQ ID NO:1 or that hybridizes to SEQ ID NO:1. Making all possible single amino acid substitutions, in an 3621 nucleotide long nucleic acid like that of SEQ ID NO:1 would require making and analyzing  $19^{3621}$  nucleic acids; these nucleic acids would have about 99.99% identity to SEQ ID NO:1. Because nucleic acids that have 90% identity to SEQ ID NO:1 would have up to 362 nucleotide substitutions, many more than  $19^{3621}$  nucleic acids would need to be made and analyzed.

Furthermore, because nucleic acids that have 90% identity to SEQ ID NO:1 would have up to 362 nucleotide substitutions, they could encode proteins with up to 362 amino acid substitutions; these proteins would have 70% identity to the 1206 amino acid long SEQ ID NO:2. The specification provides no guidance for which 362 amino acids to substitute. Thus, undue trial and error experimentation would be required to make the claimed nucleic acids.

Similarly, nucleic acids that have 93% identity to SEQ ID NO:1 would have up to 253 nucleotide substitutions, nucleic acids that have 94% identity to SEQ ID NO:1 would have up to 217 nucleotide substitutions and nucleic acids that have 90% identity to SEQ ID NO:1 would have up to 181 nucleotide substitutions, producing proteins with 79%, 82% and 85% identity, respectively, to the 1206 amino acid long SEQ ID NO:2.

Guo et al (2004, Proc. Natl. Acad. Sci. USA 101: 9205-9210) teach that while proteins are fairly tolerant to mutations resulting in single amino acid changes, increasing the number of

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substitutions additively increases the probability that the protein will be inactivated (pg 9209, right column, paragraph 2). Thus, making and analyzing proteins with up to 363 amino acid substitutions that also have pesticidal activity would require undue experimentation.

As the specification does not describe the transformation of any plant with nucleic acids with 90% identity to SEQ ID NO:1 within the full scope of the claims, undue trial and error experimentation would be required to screen through the myriad of nucleic acids encompassed by the claims and plants transformed therewith, to identify those that could control plant pests, if such plants are even obtainable.

Given the claim breadth, unpredictability in the art, undue experimentation, and lack of guidance in the specification as discussed above, the instant invention is not enabled throughout the full scope of the claims.

Applicant urges that the specification provides 14 variants of SEQ ID NO:1; these have between 38% and 92% identity across the full-length of SEQ ID NO:1 (Reply Brief pg 3-4 and Table 1).

This is not found persuasive.

First, Applicant has numerous errors in Table 1. The amino acids listed as deleted are not the deleted amino acids; they are the ones still present in the protein. Examiner has not recalculated nucleotide or amino acid identity for the sequences, but it is noted that in Applicant's previous responses, SEQ ID NO:19 was indicated as having different percent global identity to SEQ ID NO:1.



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Second, one of skill in the art would not consider a truncated protein as having 56% identity to the entire length of SEQ ID NO:2, but would only look at it as a truncated protein. One of skill in the art would only look at local match similarity. That the specification provides guidance in making truncations at amino acid 663, 699 and 667, eliminating the C-terminus, and truncations off a portion of the N-terminus (in which the first 47 amino acids are eliminated) is not disputed. However, this is not the genus of claimed nucleic acids.

Third, the specification does not consider deletions the same thing as substitutions. See pg 19, lines 3-10 and pg 28, lines 5-31, in which deletions and substitutions are discussed in the alternative.

Fourth, these sequences (except SEQ ID NO:3/4) teach a maximum of 4 amino acid substitutions. One of skill in the art would not consider these sequences as teaching which 181, 217, 253 or 362 amino acids to substitute in SEQ ID NO:2.

Fifth, SEQ ID NO:3 differs from SEQ ID NO:1 in primarily in a 400 nucleotide long region (see search results). Because the vast majority of substitutions are localized to such a small area, the protein it encodes, SEQ ID NO:4 has 89% identity to SEQ ID NO:2. Thus, SEQ ID NO:3 cannot provide guidance for making nucleic acids with 90% identity to SEQ ID NO:1 and encoding a protein with 362 amino acid substitutions relative to SEQ ID NO:2.

In summary, making fragments and a single 4 amino acid substitution are not the same thing as making 362 amino acid substitutions. The specification fails to enable nucleic acids within the full scope of the claims.

Applicant urges that SEQ ID NO:19 comprises 1860 nucleotides of SEQ ID NO:1 and retains identity, pointing to the likelihood that alteration of one or more of the 1761 additional

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nucleotides in SEQ ID NO:1 could be altered without disrupting function; furthermore, SEQ ID NO:9 encodes the same protein as SEQ ID NO:19 but has maize optimized codons, and it shares 38% global identity to SEQ ID NO:1 and 68% local identity (Reply Brief pg 4).

This is not found persuasive. SEQ ID NOs:19 and 9 encode proteins with 100% identity to the first half of SEQ ID NO:2. One of skill in the art would not consider a truncated protein as providing guidance for making amino acid substitutions in SEQ ID NO:2. Substitutions in the latter half of the protein affect the folding of the entire protein, and cannot be made willy-nilly just because a protein can be truncated.

Applicant urges that the examiner overlooks the guidance in the specification as to which 362, 253 or 217 nucleotides of SEQ ID NO:1 could be altered; guidance is provided on pg 23-24, 29 and examples 1, 6 and 7 (Reply Brief pg 8).

This is not found persuasive. Pg 23 provided general guidance for methods of making mutations in nucleic acids. It does not provide guidance for which 362 amino acid substitutions to make in SEQ ID NO:2.

Pg 24 teaches that cry proteins have conserved blocks. However, Hill et al teach that conserved blocks cannot be relied on in making amino acid substitutions (see Table 1).

The specification, on pg 29, suggests using GenBank U04365 as the other nucleic acid in shuffling to create the claimed nucleic acids; however, it encodes a protein with 79.8% identity to the instant SEQ ID NO:2. Thus, this sequence cannot be used as guidance for nucleic acids with 90% identity to SEQ ID NO:1 and that encode proteins with 70% identity to SEQ ID NO:2, as encompassed by the full scope of the claims.

Furthermore, SEQ ID NO:3 of Michaels et al is pesticidal for scarab pests (see claim 1),

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whereas the instant SEQ ID NO:1 is pesticidal to southern corn rootworm. Given the teachings of Tounsi et al, de Maagd et al, and Angsuthanasombat et al, one of skill in the art could not even predict for which pests the protein encoded by nucleic acids that have 90% identity to SEQ ID NO:1 would be toxic, and the specification does not teach which amino acid substitutions made to retain proper pesticidal activity.

Examples 1, 6 and 7 teach making the truncations and the 4 amino acid substitutions presented in Table 1 of the Reply Brief. Why these examples do not teach nucleic acids within the full scope of the claims is discussed above.

Making amino acid substitutions in *cry* proteins is unpredictable. Even a single amino acid substitution in a *cry* protein may alter its insecticidal specificity, and toxicity must be determined empirically (Tounsi et al, 2003, J. Appl. Microbiol. 95:23-28; see pg 27, column 2, paragraph 2). Thus the specification must teach how to make nucleic acids within the full scope of the claims. It does not.

The specification on pg 3, lines 4-8, even acknowledges the unpredictability in making amino acid substitutions in Cry proteins:

Although numerous investigators have attempted to make mutant endotoxin proteins with improved insecticidal activity, few have succeeded. In fact, the majority of genetically engineered *B. thuringiensis* toxins that have been reported in the literature report endotoxin activity that is no better than that of the wild-type protein, and in many cases, the activity is decreased or destroyed altogether.

Applicant urges that the examiner overlooks the guidance in Li et al, as citing on pg 25 of the specification, and Applicant's data on the truncated proteins (Reply Brief pg 8-10).

This is not found persuasive. The guidance in Li et al was not overlooked. However, Li et al only provided guidance for making truncations and insertion of chymotrypsin cleavage sites; Li et al do not provide guidance for making 181, 217, 253 or 362 amino acid substitutions

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in a 1206 amino acid protein. The instant inventors did not use Li et al to make 181, 217, 253 or 362 amino acid substitutions in a 1206 amino acid protein to create a Coleopteran pesticidal protein. Furthermore, truncated proteins are not the same as substituted proteins, as discussed above.

Applicant urges that some experimentation would be necessary to make and use other embodiments of the invention, but it would not be undue, given the guidance in the specification (Reply Brief pg 10-11).

This is not found persuasive. Such experimentation would be undue because of the lack of guidance in the specification and the unpredictability in making amino acid substitutions in *cry* proteins. See the specification on pg 3, lines 4-8, as well as Tounsi et al, de Maagd et al, and Angsuthanasombat et al, as discussed above.

Applicant urges that Lazar is drawn to a mammalian protein unrelated to the instant Cry protein, and the amino acids modified in that paper were known to be conserved, unsurprisingly leading to a loss of function (Reply Brief pg 11-12).

This is not found persuasive. Lazar et al was testing the commonly accepted notion that making conservative substitutions would not affect activity and that nonconservative substitutions would affect activity, even inactivate the protein. Lazar et al were surprised to find that this was not the case - conservative substitutions inactivated the protein and nonconservative substitutions has no effect on activity. This is relevant to the instant case because the specification, on pg 28, lines 5-16, teaches that amino acid substitutions should be conservative. Lazar et al teaches that functional substitutions must be determined empirically. The specification does teach which substitutions to make within the full scope of the claims.

Applicant urges that Hill et al is drawn to an enzyme unrelated to the instant Cry protein, and the modified residues were highly conserved, unsurprisingly leading to a loss of function (Reply Brief pg 12-13).

This is not found persuasive. Hill et al was testing the commonly accepted notion that amino acids conserved across proteins from a wide range of species can only tolerate conservative substitutions if they can tolerate any at all. Hill et al were surprised to find that this was not the case - nonconservative substitutions had no effect on activity and conservative substitutions inactivated the protein. This is relevant to the instant case because the specification, on pg 28, lines 5-16, teaches that amino acid substitutions should be conservative and on pg 24-25 teaches that other cry proteins can be used as comparisons when making the claimed nucleic acids. Hill et al teaches that functional substitutions must be determined empirically. The specification does teach which substitutions to make within the full scope of the claims.

Applicant urges that Lazar et al and Hill et al demonstrate that one of skill in the art could make substitutions in proteins and test for activity (Reply Brief pg 13).

This is not found persuasive. Lazar et al and Hill et al made single amino acid substitutions. The claims encompass nucleic acids that having 90% identity to SEQ ID NO:1 and encoding functional Cry proteins with 362 amino acid substitutions. Guo et al teaches that increasing the number of substitutions above single amino acid changes additively increases the probability that the protein will be inactivated (pg 9209, right column, paragraph 2). Thus, without further guidance, one of skill in the art could not make this many substitutions and still produce functional proteins.

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Applicant urges that there is guidance in the specification for making modifications (Reply Brief pg 13).

This is not found persuasive because the specification fails to provide guidance for making nucleic acids that having 90% identity to SEQ ID NO:1 and encoding functional Cry proteins with 362 amino acid substitutions, as encompassed by the full scope of the claims.

4. Claims 1-3, 9-12, 17-19, 38, 43, 46, 49, 52 and 55-64 are rejected under 35 U.S.C. 112, first paragraph, as containing subject matter that was not described in the specification in such a way as to reasonably convey to one skilled in the relevant art that the inventor(s), at the time the application was filed, had possession of the claimed invention. The rejection is repeated for the reasons of record as set forth in the Office action mailed 6 May 2005. Applicant's arguments filed 23 February 2006 in a Reply Brief have been fully considered but they are not persuasive.

A full review of the specification indicates that nucleic acids that have 90%, 93%, 94% or 95% identity to SEQ ID NO:1 and that encode Coleopteran-pesticides are essential to the operation of the claimed invention.

The level of skill and knowledge in the art at the time of filing was such that while large number of Coleopteran-pesticides were known, nucleic acids that have 90%, 93%, 94% or 95% identity to SEQ ID NO:1 and that encode such Coleopteran-pesticides were not.

The claim is directed to a genus of nucleic acids that have 90%, 93%, 94% or 95% identity to SEQ ID NO:1. Nucleic acids that have 90%, 93%, 94% or 95% identity to SEQ ID NO:1 would have up to 362 nucleotide substitutions, they could encode proteins with up to 362,

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253, 217 or 181 amino acid substitutions relative to the to the 1206 amino acid long SEQ ID NO:2.

The structural features that distinguish Coleopteran-pesticide-encoding nucleic acids that have 90%, 93%, 94% or 95% identity to SEQ ID NO:1 from other that have 90%, 93%, 94% or 95% identity to SEQ ID NO:1 are not described in the specification. The specification recites no structure required for Coleopteran-pesticide activity. The necessary and sufficient structural elements of a protein with Coleopteran-pesticide activity are not described.

The only species described in the specification is SEQ ID NO:1, which encodes SEQ ID NO:2, a truncated version of SEQ ID NO:1, and a few nucleic acids with a few nucleotide substitutions relative to SEQ ID NO:1.

One of skill in the art would not recognize that Applicant was in possession of the necessary common attributes or features of the genus in view of the disclosed species. Since the disclosure fails to describe the common attributes that identify members of the genus, and because the genus is highly variant, SEQ ID NO:1 alone is insufficient to describe the claimed genus.

Hence, Applicant has not, in fact, described nucleic acids with 90%, 93%, 94% or 95% identity to SEQ ID NO:1 and that encode a protein pesticidal for at least one pest belonging to the order Coleoptera within the full scope of the claims, and the specification fails to provide an adequate written description of the claimed invention.

Therefore, given the lack of written description in the specification with regard to the structural and physical characteristics of the claimed compositions, it is not clear that Applicant was in possession of the genus claimed at the time this application was filed.

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Applicant urges that the specification provides numerous nucleic acids that fall within the scope of the claims as well as much guidance for making alterations (Reply Brief pg 15).

This is not found persuasive because the structure of nucleic acids that have 90%, 93%, 94% or 95% identity to SEQ ID NO:1 and up to 362, 253, 217 or 181 amino acid substitutions, respectively, relative to the 1206 amino acid long SEQ ID NO:2 as encompassed by the full scope of the claims are not described.

Applicant urges that in *Lilly* the patent did not identify the sequence of any embodiment, while here a representative sequence is provided (Reply Brief pg 15).

This is not found persuasive. In *Lilly* the claims were drawn to the genus of vertebrate and mammalian cDNAs and the rat cDNA was described. Lilly stated at pg 1406:

... A description of a genus of cDNAs may be achieved by means of a recitation of a representative number of cDNAs, defined by nucleotide sequence, falling within the scope of the genus or of a recitation of structural features common to the members of the genus, which features constitute a substantial portion of the genus.

In the instant case, only a portion of the structural features have been described - the percent identity to SEQ ID NO:1. But because this includes nucleic acids in which the protein sequence has a very large number of amino acid substitutions, those amino acid substitutions that do not alter the function of the protein must be described. They are not. A representative number of nucleic acids that have 90%, 93%, 94% or 95% identity to SEQ ID NO:1 and up to 362, 253, 217 or 181 amino acid substitutions, respectively, relative to the 1206 amino acid long SEQ ID NO:2 are not described.

5. Claims 39-40, 44-45 and 50-51 are objected to as being dependent upon a rejected base claim, but would be allowable if rewritten in independent form including all of the limitations of the base claim and any intervening claims.



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**Conclusion**

6. No claim is allowed.

7. Any inquiry concerning this communication or earlier communications from the examiner should be directed to Anne R. Kubelik, whose telephone number is (571) 272-0801. The examiner can normally be reached Monday through Friday, 8:30 am - 5:00 pm.

If attempts to reach the examiner by telephone are unsuccessful, the examiner's supervisor, Anne Marie Grunberg, can be reached at (571) 272-0975.

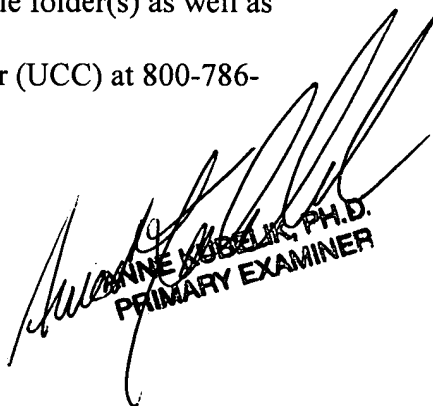
The central fax number for official correspondence is (571) 273-8300.

Any inquiry of a general nature or relating to the status of this application or proceeding should be directed to (571) 272-0547.

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Anne Kubelik, Ph.D.  
April 28, 2006



ANNE KUBELIK, PH.D.  
PRIMARY EXAMINER



ANNE MARIE GRUNBERG  
SUPERVISORY PATENT EXAMINER



Db 421 GGATTAGGTAAATACCAATTATATCTAACTGCGCTTGAAGAAATGGAAGAAATCCA 480  
Qy 481 AATGGTTCAAGAGCTTACGAGATGTCGGAATTCGAATTCGAATTCGAGATGTTATTT 540  
Db 481 AATGGTTCAAGAGCTTACGAGATGTCGGAATTCGAATTCGAATTCGAGATGTTATTT 540  
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Db 601 ACACAGGCGACCACTTCAATTTACTGTATTAAGGAGCGTCAATTTTGGAGAGAA 660  
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Qy 1561 GATAAAATCACTCAAAATTCGCGCGCTTAAATGTTGGGATAAATTTTACCGTTGT ----- 1613  
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Qy 1614 -----TTCAGTGTGTAAGGACCGAGACATACAGAGGGGGATTTATACAGTAT 1662  
Db 1621 GGGGCAATTAATACCGTTGATCGGGTCTCGGATTTACAGGGGGGGGATTAATAAGAT 1680  
Qy 1663 AATAGAGTACTGTTCTGTAGGAACCTTTATTTCTAGCTCGATATGGCTAGCATTAAGAA 1722  
Db 1681 AATAGAAATGAGTAATTAATCAATATGCGTGTAAATTTTCAAGCAATTAACAAAGAA 1740  
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Qy 1843 AAAACCTTTAAAGTTGCGATGCTATCAACAAATTAATTTAGCAACAGATAGTTGCGCTA 1902  
Db 1861 AATAAATTAATTAATGCGACTTTGCGCCCTATTAATTTTACGACAAACCGACCTTTCAAT 1920  
Qy 1903 GATTTGAAACAAATTAATTTAGGTGAAGACCTTAATTAATTAATTAATTAATTAATTAATTAAT 1962  
Db 1921 ACTCTAGGGGCTATATTTGAAGCGGAGACTTTCTTGGAAAT -----TGAAGCTTATATA 1974  
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Db 2995 GAAATACAGGAGTAACTATATACCAAGTTTACAGAAATTAACAGATCCACTCCACAGCG 3054
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Db 3595 TATATGAAAGTGTAGATTTGATTTAGACCTGAGATTA 3633
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## RESULT 4

US-10-032-717-28  
; Sequence 28, Application US/10032717  
; Patent No. US20020151709A1

## GENERAL INFORMATION:

; APPLICANT: Andre R. Abad  
; APPLICANT: Nicholas B. Duck  
; APPLICANT: Xiang Peng  
; APPLICANT: Ronald D. Flannagan  
; APPLICANT: Theodore W. Kahn  
; APPLICANT: Lynn S. Sims  
; TITLE OF INVENTION: Genes Encoding No. US20020151709A1el Proteins With  
; FILE REFERENCE: 35718/237005  
; CURRENT APPLICATION NUMBER: US/10/032.717  
; PRIORITY FILING DATE: 2001-10-23  
; PRIOR APPLICATION NUMBER: 60/242,838  
; NUMBER OF SEQ ID NOS: 48  
; SOFTWARE: FastSeq for Windows Version 4.0  
; SEQ ID NO 28  
; LENGTH: 6613  
; TYPE: DNA  
; ORGANISM: Bacillus thuringiensis  
; FEATURE:  
; NAME/KEY: misc feature  
; LOCATION: (0) - (0)  
; OTHER INFORMATION: Genomic Cyl1218-2  
US-10-032-717-28

Query Match 87.0%; Score 3151.8; DB 12; Length 6613;  
Best Local Similarity 92.1%; Pred. No. 0;  
Matches 3353; Conservative 0; Mismatches 262; Indels 24; Gaps 2;

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QY 121 GATTATAAAGATTATTTAAATAATGCTGCGGAAATGCTAGTCAATACCCCTGCTCACCT 180
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